



**International Journal of Biology, Pharmacy  
and Allied Sciences (IJBPAS)**  
*'A Bridge Between Laboratory and Reader'*

[www.ijbpas.com](http://www.ijbpas.com)

---

**EFFECT OF DIAZINON ON GONADOTROPINS AND TESTOSTERONE LEVELS  
IN SERUM OF MALE RAT**

**MOHAMMAD NABI<sup>1</sup> AND AFSHINDAVASAZ TABRIZI<sup>2\*</sup>**

**1:** Department of Veterinary Medicine, Tabriz Branch, Islamic Azad University, Tabriz, Iran

**2:** Department of Clinical Science, Collage of Veterinary Medicine, Tabriz Branch, Islamic Azad University, Tabriz, Iran

**\*Corresponding Author's: E Mail:** [afshindavasaz@yahoo.com](mailto:afshindavasaz@yahoo.com); [davasaz@iaut.ac.ir](mailto:davasaz@iaut.ac.ir)

**ABSTRACT**

Diazinon is insecticides widely used in agriculture and pest control in the environment, which can be highly toxic. DZN is bioactivated by cytochrome P450 enzymes through desulphuration to its corresponding Oxon derivative. The aim of present study was to evaluate the effect of Diazinon on gonadotropins and testosterone levels in serum of male Rat. In this study, 14 male Wistar rats were divided into 2 equal groups: group1; normal control which were received standard diet during the experiment, group 2 received Diazinon with corn oil at a dose of 75mg/kg (1.4LD50) daily besides of standard diet for 6 weeks. Diazinon was given to animals through gastric gavage. After mentioned periods, blood samples were obtained and serum was isolated through centrifuge at the 2000 RPM for 15 minutes. The mentioned parameters were measured using the available kits (Pishtazteb Ltd.) and ELISA methods. It shows that there is significant difference among groups in term of the serum value of FSH and Testosterone ( $P < 0.05$ ) but not in LH ( $p > 0.05$ ). Data showed that FSH decreased significantly in treatment group; contrary, testosterone increased. In summary, diazinon has adversely affected the reproduction organ in male rats. The present results indicate that exposure to DZN has direct effects on rat's testis and the imbalance of circulating testosterone and gonadotropins may reduce fertility.

**Keywords: Diazinon, Testosterone, LH, FSH, Rat**

---

---

## INTRODUCTION

Diazinon is a thiophosphoric acid ester developed in 1952 by Ciba-Geigy, a Swiss chemical company. It is a nonsystemic organophosphate insecticide formerly used to control cockroaches, silverfish, ants, and fleas in residential, non-food buildings. Diazinon was heavily used during the 1970s and early 1980s for general-purpose gardening use and indoor pest control. A bait form was used to control scavenger wasps in the western U.S. Diazinon is used in flea collars for domestic pets in Australia and New Zealand. Residential uses of diazinon were outlawed in the U.S. in 2004 but it is still approved for agricultural uses. An emergency antidote is atropine (Budavari, 1996).

The structure of diazinon contains a thiophosphoric ester. The phosphorus center is the reactive site of the chemical. However, no known mechanisms currently exist. A novel mechanism does exist, which proposes that the sulfur is protonated in acidic medium via a hydronium ion which ultimately delivers a hydroxide group to the phosphorus center and can react readily.

The form of diazinon varies widely as it can be in dusts, granules, liquids, concentrates, microencapsulations, wettable powders, and seed dressings. Its appearance varies depending on purity, ranging from a dark brown (industrial grade), to a colorless

liquid (pure). Indicative of its functionality of a thiophosphoric ester, the chemical has a pronounced smell similar to that of diethyl ether (Sharom *et al.*, 1980).

Diazinon is relatively stable under standard ambient temperature and pressure but is susceptible to form toxic phosphine gas upon heating. Furthermore, diazinon can oxidize into toxic phosphorus oxides if in the presence of a strong oxidizing agent.

Diazinon functions as an acetylcholinesterase (AChE) inhibitor. This enzyme breaks down the neurotransmitter acetylcholine (ACh) into [choline] and an acetate group. The inhibition of the AChE causes an abnormal accumulation of ACh in the synaptic cleft. When Diazinon enters the body, it is oxidatively degenerated to diazoxon, an organophosphate compound that is much more poisoning than diazinon and causes mainly the inhibition of AChE. The activation of diazinon is located in the liver microsomal enzyme system and requires O<sub>2</sub> and NADPH. Additionally it can also be degenerated via oxidation in the microsomal enzyme system. Both reactions are possible, and likely are catalyzed nonspecifically by the same mixed function oxidase (Hansch *et al.*, 1995; Geller *et al.*, 2003). Once formed diazoxon is biotransformed again as it is degenerated. Diazoxon is degenerated due to

the very effective hydrolyases in the microsomal and other sub cellular factions within the liver. Mammalian specie degenerate diazoxon at a much slower rate though (with a half-life of 2 to 6 weeks). Insects lack this hydrolysis step which allows for the toxic species to accumulate rapidly. The detoxification of diazoxon is processed through the mircosomal mixed function oxidase system. Although not fully known, it is believed that this is the cause for the selectivity of diazinon against insects. After the hydrolysis or oxidation diazinon is further degenerated. The aim of present study was to evaluate the effect of Diazinon on gonadotropins and testosterone levels in serum of male Rat.

#### MATERIALS AND METHODS

In this study, 14 male Wistar rats ( $200 \pm 2.1$ g and 2-3 month age) were selected for the study and were purchased from Animal House, Islamic Azad University and randomly divided into 2 equal groups: group1; normal control which were received standard diet during the experiment, group 2 received Diazinon with corn oil at a dose of 75mg/kg (1.4LD50) daily besides of standard diet for 6 weeks. Diazinon was given to animals through gastric gavage.

Animal care and experiments confirmed with the Guide for the Care and Use of Laboratory Animals of China and approval of the ethics committee of Islamic Azad University was obtained before the commencement of the study. The animals were housed under standard environmental conditions ( $23 \pm 1^\circ\text{C}$ , with  $55 \pm 5\%$  humidity and a 12 h light/12 h dark cycle) and maintained with free access to water and a standard laboratory diet *ad libitum*. After mentioned periods, blood samples were obtained and serum was isolated through centrifuge at the 2000 RPM for 15 minutes. The mentioned parameters were measured using the available kits (Pishtazteb Ltd.) and ELISA methods.

The Statistical Package for Social Sciences (SPSS Inc., Chicago, IL, USA), version 13.0, was used for statistical analysis. The data obtained were tested by ANOVA followed by T-test.  $P < 0.05$  was considered statistically significant.

#### RESULTS

Data related to measurement of FSH, LH and testosterone in both groups are presented in **Table 1**.

**Table 1: serum value of FSH, LH and testosterone in animals before and after Diazinon administration**

Group	FSH (mIU/ml)	LH (mIU/ml)	Testosterone (ng/ml)
Control	$0.2 \pm 0.02$	$1.01 \pm 0.16$	$1.22 \pm 0.06$
Treatment	$0.1 \pm 0.015$	$0.96 \pm 0.08$	$1.5 \pm 0.49$
p-value	$< 0.05$	$> 0.05$	$< 0.05$

As seen in **Table 1** it shows that there is significant difference among groups in term of the serum value of FSH and Testosterone ( $P < 0.05$ ) but not in LH ( $p > 0.05$ ). Data showed that FSH decreased significantly in treatment group; contrary, testosterone increased.

#### DISCUSSION AND CONCLUSION

Some studies have showed that DZN was capable of inducing structural and functional changes (**Wesseling et al., 1999**), and some biochemical alterations in the ovaries and testes (**Dutta et al., 2003**). Our results demonstrate that administration of Diazinon can adversely affect reproductive function by increasing the mass of testis and testosterone blood levels, but decreasing the concentrations of circulating gonadotropins.

Some histopathological studies in animals have showed that diazinon treatment may inhibit spermatogenesis and make destruction of all kind of seminiferous tubules. Maintaining LH serum levels is very important for initiating and supporting spermatogenesis, hence degeneration of Sertoli cells and germinal cells may be due to high concentrations of circulating LH (**Shan et al., 1995; Sarkar et al., 2000**). OP cause microtubule disruption of epithelium (**Nakai et al., 1995**), and finally leads to tubular atrophy (**Hess et al., 2000**) Also, spermatogenesis disruption might be due

to increase in the serum LH concentration which is detrimental to the germinal cells (having a very important role in the first steps of spermatogenesis process) (**Boekelheide and Schoenfeld, 2001; Izumi et al., 2005**).

Toxic effects of diazinon might be mediated by stimulation of lipid peroxidation, changes in the actions of antioxidant enzymes, DNA damage, and free radicals which involved in cell death (**Izumi et al., 2005; Ibrahim and El-Gamal, 2003**).

Several studies have showed increased frequencies of sperm aneuploidy observed in individuals occupationally exposed to DZN (**Narayana et al., 2006**). Also several OPs have been extensively studied as genotoxic chemicals in mammalian germinal cells and spermatids (**Recio et al., 2001; Cakir et al., 2005**).

Prolonged IP exposure to the DZN, decreased the body weight in consensus with the earlier reports, and that effect was due to diminished food and water intake in treated mice of DZN group (**Prashanthi et al., 2006**).

DZN has also been described as genotoxic, mutagenic carcinogenic agents mutagenic agent using in vivo and in vitro models (**Aluigi et al., 2005; LeBlanc et al., 1997**) probably by sharing the alkylation properties of many other OP compounds.

DZN exposure also has caused reduction in the Leydig cells.

There are several possible mechanisms for the antigonadal actions of organophosphates; they may exert a direct inhibitory action on the testis; they may affect the pituitary, causing changes in gonadotrophins concentrations and thus subsequent spermatogenic impairment; or they may change the concentration of neurotransmitter (Pidoux *et al.*, 2007).

Antiandrogens can disrupt male differentiation by several mechanisms, including antagonism of receptor binding, or by inhibition of the production, transport, or metabolism of androgens (Chattopadhyay *et al.*, 2005). The damage may have occurred by direct toxic effects of DZN on cells or tissue, but it might also occur because of imbalanced hormones levels. DZN may affect directly upon testis tissue or by entering into the pituitary gland which could cause changes of gonadotropins arise. In these actions of DZN, serum LH and FSH levels increased probably due to suppression of feed-back inhibition of anterior pituitary. The suppression of feedback inhibition may secondarily increase the LH and FSH secretion. The results of the present study indicate that DZN has a direct effect on pituitary, which lead to increase in circulating LH levels. Diazinon increase serum FSH levels after

injection in mice. FSH has important effects on sertoli cells. Inhibin and other factors secreted by Sertoli cells cause increase of circulating FSH levels by feedback on pituitary. Increase of LH levels has toxic effects on testis tissue.

In summary, diazinon has adversely affected the reproduction organ in male rats. The present results indicate that exposure to DZN has direct effects on rat's testis and the imbalance of circulating testosterone and gonadotropins may reduce fertility.

## REFERENCES

- [1] Aluigi MG, Angelini C, Falugi C, Fossa R, Genever P, Gallus L, et al. Interaction between organophosphate compounds and cholinergic functions during development. *ChemBiol Interact* 2005; 157-158:305-316.
- [2] Boekelheide K, Schoenfeld HA. Spermatogenesis by Sisyphus: proliferating stem germ cells fail to repopulate the testis after 'irreversible' injury. *AdvExp Med Biol* 2001; 500: 421-428.
- [3] Budavari, S. (1996). *The Merck Index - An Encyclopedia of Chemicals, Drugs, and Biologicals*. Whitehouse Station, NJ: Merck. p. 508.
- [4] Cakir S, Sarikaya R. Genotoxicity testing of some organophosphate

- insecticides in the *Drosophila* wing spot test. *Food Chem Toxicol* 2005; 43: 443-450.
- [5] Chattopadhyay A, Sarkar M, Biswas NM. Dose-dependent effect of copper chloride on male reproductive function in immature rats. *Kathmandu Univ Med J (KUMJ)* 2005; 3: 392-400.
- [6] Civen M, Brown CB. The effect of organophosphate insecticides on adrenal corticosterone formation. *Pesticide Biochemistry and Physiology* 1974; 4: 254–259.
- [7] Dutta HM, Meijer HJ. Sublethal effects of diazinon on the structure of the testis of bluegill, *Lepomis macrochirus*: a microscopic analysis. *Environmental pollution* 2003; 125:355-360.
- [8] Geller, Robert J.; Lopez, Gaylord P.; Cutler, Stephen; Lin, Diana; Bachman, George F.; Gorman, Susan E. (2003). Atropine availability as an antidote for nerve agent casualties: Validated rapid reformulation of high-concentration atropine from bulk powder. *Annals of Emergency Medicine* 41 (4): 453–6.
- [9] Hansch, Corwin; Leo, Albert; Hoekman, David (1995). *Exploring QSAR: Volume 2: Hydrophobic, Electronic, and Steric Constants*. Washington, DC: American Chemical Society. p. 106. ISBN 978-0-8412-2991-4.
- [10] Hatjian BA, Mutch E, Williams FM, Blain PG, Edwards JW. Cytogenetic response without changes in peripheral cholinesterase enzymes following exposure to a sheep dip containing diazinon in vivo and in vitro. *Mutat Res* 2000; 427, 85-92.
- [11] Hess RA, Nakai M. Histopathology of the male reproductive system induced by the fungicide benomyl. *Histol Histopathol* 2000; 15: 207-224.
- [12] Ibrahim NA, El-Gamal BA. Effect of diazinon, an organophosphate insecticide, on plasma lipid constituents in experimental animals. *J Biochem Mol Biol* 2003; 36: 499-504.
- [13] Izumi H, Kimura E, Ota T, Shimazu S. A two-generation reproductive toxicity study of n-butylbenzene in rats. *J Toxicol Sci* 2005; 30: 21-38.
- [14] LeBlanc G, Bain LG, Wilson VS. Pesticides: Multiple mechanisms of demasculinization. *Mol Cell Endocrinol* 1997; 126:1-5.
- [15] Maitra SK, Sarkar R. Influence of methyl parathion on gametogenic

- and acetylcholinesterase activity in the testis of whitethroatedmunia (*Lonchuramalabarica*). Arch Environ Contam Toxicol 1996; 30: 384-389
- [16] Nakai M, Hess RA, Netsu J, Nasu T. Deformation of the rat Sertoli cell by oral administration of carbendazim (methyl 2-benzimidazole carbamate). J Androl 1995; 16: 410-416.
- [17] Narayana K, Prashanthi N, Nayanatara A, Bairy LK, D'Souza UJ. An organophosphate insecticide methyl parathion (o- o- dimethyl o-4 - nitrophenylphosphorothioate) induces cytotoxic damage and tubular atrophy in the testis despite elevated testosterone level in the rat. J ToxicolSci 2006; 31:177-189.
- [18] Pidoux G, Gerbaud P, Tsatsaris V, Marpeau O, Ferreira F, Meduri G, et al. Biochemical characterization and modulation of LH/CG-receptor during human trophoblast differentiation. J Cell Physiol 2007; 212: 26-35.
- [19] Prashanthi N, Narayana K, Nayanatara A, Chandra Kumar HH, Bairy KL, D'Souza UJ. The reproductive toxicity of the organophosphate pesticide 0, 0-dimethyl 0-4-nitrophenyl phosphorothioate (methyl parathion) in the male rat. Folia Morphol (Warsz) 2006; 65: 309-321.
- [20] Recio R, Robins WA, Ocmpongomez G, Borja-Aburto V. Organophosphorus pesticide exposure increase the frequency of sperm sex null aneuploidy. Environ Health Prspect 2001; 109:1237-1240.
- [21] Salem KA. Infectious diseases among food handlers. J Egypt Public Health Assoc 1998; 73: 563-575.
- [22] Sarkar R, Mohanakumar KP, Chowdhury M. Effects of an organophosphate pesticide, quinalphos, on the hypothalamo-pituitary-gonadal axis in adult male rats. Journal of Reproduction and Fertility 2000; 118: 29-38
- [23] Shan L, Hardy DO, Catterall JF, Hardy MP. Effects of luteinizing hormone (LH) and androgen on steady state levels of messenger ribonucleic acid for LH receptors, androgen receptors, and steroidogenic enzymes in rat Leydig cell progenitors in vivo. Endocrinology 1995; 136: 1686-1693.
- [24] Sharom, M.S.; Miles, J.R.W.; Harris, C.R.; McEwen, F.L.

(1980).Behaviour of 12 insecticides in soil and aqueous suspensions of soil and sediment. *Water Research* 14 (8): 1095–100.

[25] Watanabe HK, Hoskins B, Ho IK. Selective inhibitory effect of organophosphates on UDP-glucuronyltransferase activities in rat liver microsomes. *BiochemPharmacol* 1986; 35:455-460.

[26] Wesseling C, Antich D, Hogstedt C, Rodríguez AC, Ahlbom A. Geographical differences of cancer incidence in Costa Rica in relation to environmental and occupational pesticide exposure. *Int J Epidemiol* 1999; 28: 365-374.